

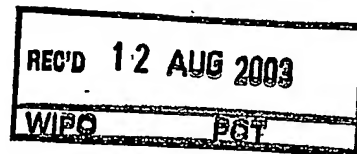


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If no title is shown please refer to the description.  
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Camp-regulated phosphorprotein for diagnostic and therapeutic use in  
neurodegenerative diseases

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## **cAMP-REGULATED PHOSPHOPROTEIN FOR DIAGNOSTIC AND THERAPEUTIC USE IN NEURODEGENERATIVE DISEASES**

The present invention relates to methods of diagnosing, prognosticating and monitoring the progression of neurodegenerative diseases in a subject. Furthermore, methods of therapy control and screening for modulating agents of neurodegenerative diseases are provided. The invention also discloses pharmaceutical compositions, kits, and recombinant animal models.

Neurodegenerative diseases, in particular Alzheimer's disease (AD), have a strongly debilitating impact on a patient's life. Furthermore, these diseases constitute an enormous health, social and economic burden. Alzheimer's disease is the most common age-related neurodegenerative condition affecting about 10 % of the population over 65 years of age and up to 45 % over age 85 (for a recent review see Vickers et al., *Progress in Neurobiology* 2000, 60: 139-165). Presently this amounts to an estimated 12 million cases in the US, Europe, and Japan. This situation will inevitably worsen with the demographic increase in the number of old people ("aging of the baby boomers") in developed countries. The neuropathological hallmarks that occur in the brains of individuals with Alzheimer's disease are senile plaques, composed of amyloid- $\beta$  protein, and profound cytoskeletal changes coinciding with the appearance of abnormal filamentous structures and the formation of neurofibrillary tangles. AD is a progressive disease that is associated with early deficits in memory formation and ultimately leads to the complete erosion of higher cognitive function. A characteristic feature of the pathogenesis of AD is the selective vulnerability of particular brain regions and subpopulations of nerve cells to the degenerative process. Specifically, the temporal lobe region and the hippocampus are affected early and more severely during the progression of the disease. On the other hand, neurons within the frontal cortex, occipital cortex, and the cerebellum remain largely intact and are protected from neurodegeneration (Terry et al., *Annals of Neurology* 1981, 10: 184-192).

Currently, there is no cure for AD, nor is there an effective treatment to halt the progression of AD or even to diagnose AD ante-mortem with high probability. Several risk factors have been identified that predispose an individual to develop AD, among them most prominently the epsilon4 allele of apolipoprotein E (ApoE). Although there are rare examples of early-onset AD which have been attributed to genetic defects in the genes for amyloid precursor protein (APP), presenilin-1, and presenilin-2, the prevalent form of late-onset sporadic AD is of hitherto unknown etiologic origin. The late onset and complex pathogenesis of neurodegenerative disorders pose a formidable challenge to the development of therapeutic and diagnostic agents.

It is crucial to expand the pool of potential drug targets and diagnostic markers. It is therefore an object of the present invention to elucidate the causes and the underlying pathogenesis of neurological diseases and to provide methods, materials, and animal models which are suited inter alia for the diagnosis and development of a treatment of these diseases. This object has been solved by the features of the independent claims. The subclaims define preferred embodiments of the present invention.

A group of cAMP-regulated phosphoproteins (ARPPs) has been shown to function as so called intracellular third-messengers in the mammalian central nervous system. Receptor-mediated phosphorylation and dephosphorylation of ARPPs constitute important pathways for the regulation of neuronal functions in response to levels of the important second messenger cAMP and activity of the cAMP-dependent protein kinase, PKA (for recent review, Greengard, Science 2001, 294: 1024-1030). To date, the best characterized ARPPs are DARPP-32 (dopamine and cAMP regulated phosphoprotein of 32 kDa molecular weight), ARPP-16/19 (cAMP regulated phosphoprotein of 16/19 kDa molecular weight), and ARPP-21 (cAMP regulated phosphoprotein of 21 kDa molecular weight), all of which are encoded by separate genes in the human genome. Thus, DARPP-32 is encoded on chromosome 17, ARPP-16/19 on chromosome 15, and the ARPP-21 locus is found on chromosome 3 of the human genome. DARPP-32, ARPP-16/19, and ARPP-21 are non-

homologous proteins but may have similar or even overlapping functions based on their tissue expression pattern within the human post-mortem brain. Using in situ hybridization techniques, transcripts for all three ARPPs can be detected in brain regions that receive a rich dopamine innervation from the mesencephalon, i.e. the caudate nucleus, putamen, nucleus accumbens, and the amygdaloid complex. ARPP-16/19, in addition, shows a strong mRNA hybridization signal in the neocortex, whereas DARPP-32 and ARPP-21 showed low levels of signal intensity only (Brene et al., J Neurosci 1994, 14: 985-998). The distribution of ARPP mRNAs overlaps to a large extent with the distribution of the dopamine D1 receptor which thus may regulate the phosphorylation status of ARPPs via adenylate cyclase/cAMP and PKA. In fact, the phosphorylation status of DARPP-32 is at the crossroads of multiple complex signaling pathways involving PKA (signaling by receptors for dopamine, opiate, adenosine, serotonin, vasoactive intestinal peptide), the protein phosphatase PP-2B/calcineurin (signaling by receptors for dopamine, gamma-aminobutyric acid, glutamate), and the protein phosphatase PP-1 which controls the state of phosphorylation and activity of numerous physiologically important substrates including neurotransmitter receptors, voltage-gated ion channels, ion pumps, and transcription factors (Greengard, Science 2001, 294: 1024-1030).

A function of ARPP-21 is largely unknown. Human ARPP-21 consists of 89 amino acids and is phosphorylated by PKA on Serin-56 (Brene et al., J Neurosci 1994, 14: 985-998). The human ARPP-21 isoform cARPP encoding a polypeptide of 89 amino acids has been described in WO00/34477. Available evidence supports the view that ARPP-21 is a cAMP regulated phosphoprotein highly enriched in the cell bodies and terminals of medium-sized spiny neurons of the basal ganglia with the highest levels of immunoreactivity seen in structures comprising the limbic striatum (Ouimet et al., J Neurosci 1989, 9: 865-875). ARPP-21 may therefore play a role as an intracellular third messenger in mediating some of the effects of dopamine, vasoactive intestinal polypeptide, and/or other neurotransmitters acting via cAMP in these dopamine-innervated brain regions (Ouimet et al., J Neurosci 1989, 9: 865-875; Hemmings and Greengard, J Neurosci 1989, 9: 851-864).

In fact, the dopamine D1 agonist SKF38393 was shown to increase the state of phosphorylation of ARPP-21 in tissue slices of the substantia nigra of rat brain (Tsou et al., J Neurochem 1993, 60: 1043-1046). Using mouse striatal slices these results were recently corroborated and extended to show that agonists of dopamine D2 receptors cause a strong decrease in ARPP-21 phosphorylation (Caporaso et al., Neuropharmacology 2000, 39: 1637-1644). The likely effector of the dopamine D2 receptor signal is the protein phosphatase PP-2A. Several neurological and psychiatric diseases are associated with abnormalities in the dopamine signaling pathways, among them Parkinson's disease, schizophrenia, attention deficit hyperactivity disorder, and drug abuse. A dysregulation of DARPP-32 function has been postulated to be causally related to the above disorders and, therefore, DARPP-32 can be considered a potential therapeutic target for said diseases (WO 99/20273; US 5777195). A recent study correlates levels of the cAMP regulated phosphoprotein ARPP-19 mRNA and protein in brain tissue from patients suffering from Down syndrome or from Alzheimer's disease (Kim et al., J Neural Transm Suppl 2001, 61: 263-272). Kim and coworkers find normal levels of ARPP-19 mRNA in the temporal lobe and a reduced level of ARPP-19 protein in the cerebellum of AD brain tissue when compared to normal brain.

The present disclosure provides a defined pathophysiological implication and diagnostic and therapeutic utility for a novel and hitherto undescribed human isoform of the cAMP regulated phosphoprotein ARPP-21, herein designated as human TARPP (hTARPP), on the basis of a differential expression of hTARPP mRNA in post-mortem brains of patients suffering from Alzheimer's disease in comparison to age-matched healthy individuals. In the mouse, a homologous ARPP-21 splice-variant, called TARPP, encodes a ca. 100 kDa protein that accompanies T cell receptor gene rearrangement and thymocyte education (Kisielow et al., Eur J Immunol 2001, 31: 1141-1149). The name "TARPP" was coined to reflect the thymocyte-specific protein expression in mice. However, murine TARPP mRNA and protein can also be detected in the

brain, whereas no mRNA or protein is found in heart, lung, liver, lymph nodes, and spleen. A function for murine TARPP in the brain has not been described.

To date, no experiments have been described that demonstrate a link between the dysregulation of ARPP-21 gene expression and neurodegenerative disorders. Particularly the disclosure in the present invention of the novel human TARPP (hTARPP) isoform, and the identification of a link of this isoform with neurodegenerative diseases, offer new ways, inter alia, for the diagnosis and treatment of neurodegenerative disorders, in particular Alzheimer's disease.

The singular forms "a", "an", and "the" as used herein and in the claims include plural reference unless the context dictates otherwise. For example "a cell" means as well a plurality of cells, and so forth. The term "and/or" as used in the present specification and in the claims implies that the phrases before and after this term are to be considered either as alternatives or in combination. For instance, the wording "determination of a level and/or an activity" means that either only a level, or only an activity, or both a level and an activity are determined. The term "level" as used herein is meant to comprise a gage of, or a measure of the amount of, or a concentration of a transcription product, for instance an mRNA, or a translation product, for instance a protein or polypeptide. The term "activity" as used herein shall be understood as a measure for the ability of a transcription product or a translation product to produce a biological effect or a measure for a level of biologically active molecules. The term "activity" also refers to enzymatic activity. The terms "level" and/or "activity" further refer to gene expression levels or gene activity. Gene expression can be defined as the utilization of the information contained in a gene by transcription and translation leading to the production of a gene product. A gene product comprises either RNA or protein and is the result of expression of a gene. The amount of a gene product can be used to measure how active a gene is. The term "gene" as used in the present specification and in the claims comprises both coding regions (exons) as well as non-coding regions (e.g. non-coding regulatory

elements such as promoters or enhancers, introns, leader and trailer sequences). "Regulatory elements" as used in the present disclosure may include inducible and non-inducible promoters, enhancers, operators and other elements that drive and regulate gene expression. The term "fragment" as used herein is meant to comprise e.g. an alternatively spliced, or truncated, or otherwise cleaved transcription product or translation product. The term "derivative" as used herein refers to a mutant, or an RNA-edited, or a chemically modified, or otherwise altered transcription product, or to a mutant, or chemically modified, or otherwise altered translation product. For instance, a "derivative" may be generated by processes such as altered phosphorylation, or glycosylation, acetylation or lipidation, or by altered signal peptide cleavage or other types of maturation cleavage. These processes may occur post-translationally. The term "modulator" as used in the present invention and in the claims refers to a molecule capable of changing or altering the level and/or the activity of a gene, or a transcription product of a gene, or a translation product of a gene. Preferably, a "modulator" is capable of changing or altering the biological activity of a transcription product or a translation product of a gene. Said modulation, for instance, may be an increase or a decrease in enzyme activity, a change in binding characteristics, or any other change or alteration in the biological, functional, or immunological properties of said translation product of a gene.

Neurodegenerative diseases or disorders according to the present invention comprise Alzheimer's disease, Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis, Pick's disease, fronto-temporal dementia, progressive nuclear palsy, corticobasal degeneration, cerebro-vascular dementia, multiple system atrophy, and mild-cognitive impairment. Further conditions involving neurodegenerative processes are, for instance, ischemic stroke and age-related macular degeneration.

The invention features a novel isolated nucleic acid molecule encoding a protein molecule whose amino acid sequence comprises the sequence shown in SEQ ID NO. 1. Hereinafter, the protein molecule of SEQ ID NO. 1 is



denoted human TARPP (hTARPP). Subject to the protein modules of SEQ ID NO. 1, i.e. putative consensus binding motifs for PLC $\gamma$  and Grb2, as well as an R3H domain, human TARPP may function as a cAMP regulated protein, as an intracellular third messenger, or as a scaffolding protein. Human TARPP may interact with lipids and other proteins, or it may be implicated in nucleic acid binding, in nerve cell signaling pathways, and in organizing and regulating neuronal function, and thus hTARPP may play a role in neurodegeneration, in cell protection and regeneration processes. The present invention also features functional variants, derivatives and fragments of hTARPP, which might have a modification of the given primary structure of hTARPP, but whose essential biological function may remain unaffected.

The term "variant" as used herein refers to any polypeptide and protein, compared to the polypeptides, proteins used in the present invention, in which one or more amino acids are added and/or substituted and/or deleted and/or inserted at the N-terminus or C-terminus or within the native amino acid sequences of the native polypeptides, proteins of the present invention. Additionally, the term "variant" shall include any shorter version of the polypeptides and proteins herein. A variant shall also comprise a sequence, that has at least about 85% sequence identity, more preferably at least about 90% sequence identity, and most preferably at least about 95% sequence identity with the amino acid sequences of SEQ ID NO. 1. Derivatives, variants and fragments of hTARPP may include, but are not limited to functional consensus binding motifs for PLC $\gamma$  and Grb2, as well as an R3H domain or other functional modules within the polypeptide sequence of hTARPP. Variants of a protein molecule shown in SEQ ID NO. 1 include, for example, proteins with conservative amino acid substitutions in highly conservative regions. For example, isoleucine, valine and leucine can each be substituted for one another. Aspartate and glutamate can be substituted for each other. Glutamine and asparagine can be substituted for each other. Serine and threonine can be substituted for each other. Amino acid substitutions in less conservative regions include, for example, isoleucine, valine and leucine, which can each be substituted for one another. Aspartate and glutamate can

be substituted for each other. Glutamine and asparagine can be substituted for each other. Serine and threonine can be substituted for each other. Glycine and alanine can be substituted for each other. Alanine and valine can be substituted for each other. Methionine can be substituted for each of leucine, isoleucine or valine, and vice versa. Lysine and arginine can be substituted for each other. One of aspartate and glutamate can be substituted for one of arginine or lysine, and vice versa. Histidine can be substituted for arginine or lysine, and vice versa. Glutamine and glutamate can be substituted for each other. Asparagine and aspartate can be substituted for each other. Other examples of protein modifications include glycosylation and further post-translational modifications. Proteins and polypeptides of the present invention include variants, fragments and chemical derivatives of the protein comprising SEQ ID NO. 1. They can include proteins and polypeptides which can be isolated from nature or be produced by recombinant and/or synthetic means. Native proteins or polypeptides refer to naturally-occurring truncated or secreted forms, naturally occurring variant forms (e.g. splice-variants) and naturally occurring allelic variants. As used herein, protein and polypeptide refer to a linear series of amino acid residues connected to one another by peptide bonds between the alpha-amino group and carboxy groups of adjacent amino acid residues. Other covalent bonds, such as amide and disulfide bonds, may also be present.

The invention also features the nucleic acid molecules encoding functional variants of the protein molecule of SEQ ID NO. 1. Nucleic acid molecules can be DNA molecules, such as genomic DNA molecules or cDNA molecules, or RNA molecules, such as mRNA molecules. In particular, said nucleic acid molecules can be cDNA molecules comprising a nucleotide sequence of SEQ ID NO. 2 or SEQ ID NO. 3.

The invention also features an isolated DNA molecule capable of hybridizing with the complement of the cDNA described in SEQ ID NO. 2 or SEQ ID NO. 3 under stringent conditions. Stringent conditions means that hybridization will be carried out 5°C to 10°C below that temperature at which totally

complementary nucleic acids will just hybridize. Optimized stringency conditions for each sequence are established on parameters such as temperature, nucleic acid molecule consistency, salt conditions, and others well known to those of ordinary skill in the art (see Sambrook and Russell, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 2000). Examples for stringent conditions include (i) 0.2xSSC (standard saline citrate) and 0.1 % SDS at 60 °C and (ii) 50 % formamide, 4xSSC, 50 mM HEPES, pH 7.0, 10x Denhardt's solution, 100 µg/ml thermally denatured salmon sperm DNA at 42 °C. This shall not exclude even higher stringency conditions as mentioned, nor shall it exclude lower stringency conditions as mentioned.

In another aspect, the invention features a vector comprising a nucleic acid encoding a protein molecule shown in SEQ ID NO. 1. In preferred embodiments, a virus, a bacteriophage, or a plasmid comprises the described nucleic acid. In particular, a plasmid adapted for expression in a bacterial cell comprises said nucleic acid molecule, encoding a protein molecule shown in SEQ ID NO. 1, or a fragment, or a variant thereof, and the regulatory elements necessary for expression of said molecule in a bacterial cell. In a further aspect, the invention features a plasmid adapted for expression in a yeast cell which comprises a nucleic acid molecule encoding a protein molecule shown in SEQ ID NO. 1, or a variant or a fragment thereof, and the regulatory elements necessary for expression of said molecule in a yeast cell. In another aspect, the invention features a plasmid adapted for expression in a mammalian cell which comprises a nucleic acid molecule, encoding a protein molecule shown in SEQ ID NO. 1, or a fragment or a variant thereof, and the regulatory elements necessary for expression of said molecule in a mammalian cell.

In a further aspect, the invention features a cell comprising a nucleic acid molecule encoding a protein molecule shown in SEQ ID NO. 1, or a fragment or a variant thereof. The present invention also features cells comprising a DNA molecule capable of hybridizing with the complement of the c D N A

described in SEQ ID NO. 2 or SEQ ID NO. 3 under stringent conditions. In preferred embodiments, said cell is a bacterial cell, a yeast cell, a mammalian cell, or a cell of an insect. In particular, the invention features a bacterial cell comprising a plasmid adapted for expression in a bacterial cell, said plasmid comprising a nucleic acid molecule encoding a protein molecule shown in SEQ ID NO. 1, or a fragment, or a variant thereof, and the regulatory elements necessary for expression of said molecule in the bacterial cell. The invention also features a yeast cell comprising a plasmid adapted for expression in a yeast cell, said plasmid comprises a nucleic acid molecule encoding a protein molecule shown in SEQ ID NO. 1, or a fragment, or a variant thereof, and the regulatory elements necessary for expression of said molecule in the yeast cell. The invention further features a mammalian cell comprising a plasmid adapted for expression in a mammalian cell, said plasmid comprising a nucleic acid molecule encoding a protein molecule shown in SEQ ID NO. 1, or a variant, or a fragment thereof, and the regulatory elements necessary for expression of said molecule in the mammalian cell.

In one aspect the present invention features a protein molecule shown in SEQ ID NO. 1.

The invention further features an antibody specifically immunoreactive with an immunogen, wherein said immunogen is a translation product of the human TARPP gene shown in SEQ ID NO. 1, or a fragment, a variant, or a derivative thereof. The immunogen may comprise immunogenic or antigenic epitopes or portions of a translation product of said gene, wherein said immunogenic or antigenic portion of a translation product is a polypeptide, and wherein said polypeptide elicits an antibody response in an animal, and wherein said polypeptide is immunospecifically bound by said antibody. Methods for generating antibodies are well known in the art (see Harlow et al., *Antibodies, A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York). The term "antibody", as employed in the present invention, encompasses all forms of antibodies known in the art, such as

polyclonal, monoclonal, chimeric, recombinatorial, anti-idiotypic, humanized or single chain antibodies, as well as fragments thereof. Antibodies of the present invention are useful, for instance, in a variety of diagnostic and therapeutic methods involving detecting hTARPP translation products.

In a preferred embodiment of the present invention, said antibodies can be used for detecting the pathological state of a cell in a sample from a subject, comprising immunocytochemical staining of said cell with said antibody, wherein an altered degree of staining, or an altered staining pattern in said cell compared to a cell representing a known health status indicates a pathological state of said cell. The invention is particularly suited to detect pathological structures in the brain of a subject. It is also especially suited to detect pathological cells of the muscular system, prostate, stomach, testis, ovary, adrenal glands, mammary glands, liver, spleen, lung, trachea or placenta. Preferably, the pathological state relates to a neurodegenerative disease, in particular to Alzheimer's disease. Immunocytochemical staining of a cell can be carried out by a number of different experimental methods well known in the art. It might be preferred, however, to apply an automated method for the detection of antibody binding, wherein the determination of the degree of staining of a cell, or the determination of the cellular or subcellular staining pattern of a cell, or the topological distribution of an antigen on the cell surface or among organelles and other subcellular structures within the cell, are carried out according to the method described in US patent 6150173.

In one aspect, the invention features a method of diagnosing or prognosticating a neurodegenerative disease in a subject, or determining whether a subject is at increased risk of developing said disease. The method comprises: determining a level, or an activity, or both said level and said activity of (i) a transcription product of the gene coding for hTARPP, and/or of (ii) a translation product of the gene coding for hTARPP, and/or of (iii) a fragment or derivative of said transcription or translation product in a sample from said subject and comparing said level, and/or said activity to a reference

value representing a known disease or health status, thereby diagnosing or prognosticating said neurodegenerative disease in said subject, or determining whether said subject is at increased risk of developing said neurodegenerative disease.

In a further aspect, the invention features a method of monitoring the progression of a neurodegenerative disease in a subject. A level, or an activity, or both said level and said activity, of (i) a transcription product of the gene coding for hTARPP, and/or of (ii) a translation product of the gene coding for hTARPP, and/or of (iii) a fragment or derivative of said transcription or translation product in a sample from said subject is determined. Said level and/or said activity is compared to a reference value representing a known disease or health status. Thereby the progression of said neurodegenerative disease in said subject is monitored.

In still a further aspect, the invention features a method of evaluating a treatment for a neurodegenerative disease, comprising determining a level, or an activity, or both said level and said activity of (i) a transcription product of the gene coding for hTARPP, and/or of (ii) a translation product of the gene coding for hTARPP, and/or of (iii) a fragment or derivative of said transcription or translation product in a sample obtained from a subject being treated for said disease. Said level, or said activity, or both said level and said activity are compared to a reference value representing a known disease or health status, thereby evaluating the treatment for said neurodegenerative disease.

In a preferred embodiment, said subjects suffer from Alzheimer's disease. Further examples of neurodegenerative diseases are Parkinson's disease, Huntington's disease, amyotrophic lateral sclerosis, Pick's disease, fronto-temporal dementia, progressive nuclear palsy, corticobasal degeneration, cerebro-vascular dementia, multiple system atrophy, and mild-cognitive impairment. Further conditions involving neurodegenerative processes are, for instance, ischemic stroke and age-related macular degeneration.

The present invention discloses the differential expression and regulation of hTARPP in specific brain regions of Alzheimer's disease patients. Consequently, the gene coding for hTARPP and its corresponding translation products may have a causative role in the regional selective neuronal degeneration typically observed in Alzheimer's disease. Alternatively, hTARPP may confer a neuroprotective function to the remaining surviving nerve cells. Based on these disclosures, the present invention has utility for the diagnostic evaluation and prognosis as well as for the identification of a predisposition to a neurodegenerative disease, in particular Alzheimer's disease. Furthermore, the present invention provides methods for the diagnostic monitoring of patients undergoing treatment for such a disease.

It is particularly preferred that said sample to be analyzed and determined is selected from the group consisting of a brain tissue, or other tissues, organs, or body cells. The sample can also consist of cerebrospinal fluid or other body fluids including saliva, urine, blood, serum plasma, or nasal mucosa.

In further preferred embodiments, said reference value is that of a level, or an activity, or both said level and said activity of (i) a transcription product of the gene coding for hTARPP, and/or of (ii) a translation product of the gene coding for hTARPP, and/or of (iii) a fragment or derivative of said transcription or translation product in a sample from a subject not suffering from said neurodegenerative disease.

In preferred embodiments, an increase or decrease in hTARPP mRNA and /or hTARPP protein in a sample cell or tissue from said subject relative to a reference value representing a known health status indicates a diagnosis, or prognosis, or increased risk of becoming diseased with a neurodegenerative disease, particularly Alzheimer's disease.

In preferred embodiments, measurement of the level of transcription products of the gene coding for hTARPP is performed in a sample from a subject using a quantitative PCR-analysis with primer combinations to amplify said gene

specific sequences from cDNA obtained by reverse transcription of RNA extracted from a sample of a subject. A Northern blot with probes specific for said gene can also be applied. It might also be preferred to measure transcription products by means of chip-based micro-array technologies. These techniques are known to those of ordinary skill in the art (see Sambrook and Russell, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 2000).

Furthermore, a level and/or an activity of a translation product of the gene coding for hTARPP and/or a fragment of said translation product, can be detected using an immunoassay, an activity assay, and/or binding assay. These assays can measure the amount of binding between said protein molecule and an anti-protein antibody by the use of enzymatic, chromodynamic, radioactive, magnetic, or luminescent labels which are attached to either the anti-protein antibody or a secondary antibody which binds the anti-protein antibody. In addition, other high affinity ligands may be used. Immunoassays which can be used include e.g. ELISAs, Western blots and other techniques known to those of ordinary skill in the art (see Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1999). All these detection techniques may also be employed in the format of micro-arrays, protein-arrays, or protein-chip based technologies.

In a preferred embodiment, the level, or the activity, or both said level and said activity of (i) a transcription product of the gene coding for hTARPP, and/or of (ii) a translation product of the gene coding for hTARPP, and/or of (iii) a fragment or derivative of said transcription or translation product in a series of samples taken from said subject over a period of time is compared, in order to monitor the progression of said disease. In further preferred embodiments, said subject receives a treatment prior to one or more of said sample gatherings. In yet another preferred embodiment, said level and/or activity is determined before and after said treatment of said subject.



In another aspect, the invention features a kit for diagnosing or prognosticating neurodegenerative diseases, in particular Alzheimer's disease, or determining the propensity or predisposition of a subject to develop a neurodegenerative disease, in particular Alzheimer's disease, said kit comprising:

- (a) at least one reagent which is selected from the group consisting of (i) reagents that selectively detect a transcription product of the gene coding for hTARPP, (ii) reagents that selectively detect a translation product of the gene coding for hTARPP; and
- (b) instruction for diagnosing, or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, or determining the propensity or predisposition of a subject to develop such a disease by
  - detecting a level, or an activity, or both said level and said activity, of said transcription product and/or said translation product of the gene coding for hTARPP, in a sample from said subject; and
  - diagnosing or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, or determining the propensity or predisposition of said subject to develop such a disease,

wherein a varied level, or activity, or both said level and said activity, of said transcription product and/or said translation product compared to a reference value representing a known health status; or a level, or activity, or both said level and said activity, of said transcription product and/or said translation product similar or equal to a reference value representing a known disease status, indicates a diagnosis or prognosis of a neurodegenerative disease, in particular Alzheimer's disease, or an increased propensity or predisposition of developing such a disease. The kit, according to the present invention, may be particularly useful for the identification of individuals that are at risk of developing a neurodegenerative disease, in particular Alzheimer's disease. Consequently, the kit, according to the invention, may serve as a means for targeting identified individuals for early preventive measures or therapeutic intervention prior to disease onset, before irreversible damage in the course of the disease has been inflicted. Furthermore, in preferred embodiments, the kit featured in the invention is useful for monitoring a progression of a

neurodegenerative disease, in particular Alzheimer's disease, in a subject, as well as monitoring success or failure of therapeutic treatment for such a disease of said subject.

In another aspect, the invention features a method of treating or preventing a neurodegenerative disease, in particular Alzheimer's disease, in a subject comprising the administration to said subject in a therapeutically or prophylactically effective amount of an agent or agents which directly or indirectly affect a level, or an activity, or both said level and said activity, of (i) the gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii). Said agent may comprise a small molecule, or it may also comprise a peptide, an oligopeptide, or a polypeptide. Said peptide, oligopeptide, or polypeptide may comprise an amino acid sequence shown in SEQ ID NO. 1, or a fragment, or derivative, or a variant thereof.

In preferred embodiments, the method comprises the application of per se known methods of gene therapy and/or antisense nucleic acid technology to administer said agent or agents. In general, gene therapy includes several approaches: molecular replacement of a mutated gene, addition of a new gene resulting in the synthesis of a therapeutic protein, and modulation of endogenous cellular gene expression by recombinant expression methods or by drugs. Gene-transfer techniques are described in detail (see e.g. Behr, *Acc Chem Res* 1993, 26: 274-278 and Mulligan, *Science* 1993, 260: 926-931) and include direct gene-transfer techniques such as mechanical microinjection of DNA into a cell as well as indirect techniques employing biological vectors (like recombinant viruses, especially retroviruses) or model liposomes, or techniques based on transfection with DNA coprecipitation with polycations, cell membrane perturbation by chemical (solvents, detergents, polymers, enzymes) or physical means (mechanic, osmotic, thermic, electric shocks). The postnatal gene transfer into the central nervous system has been described in detail (see e.g. Wolff, *Curr Opin Neurobiol* 1993, 3: 743-748).

In particular, the invention features a method of treating or preventing a neurodegenerative disease by means of antisense nucleic acid therapy, i.e. the down-regulation of an inappropriately expressed or defective gene by the introduction of antisense nucleic acids or derivatives thereof into certain critical cells (see e.g. Gillespie, *DN&P* 1992, 5: 389-395; Agrawal and Akhtar, *Trends Biotechnol* 1995, 13: 197-199; Crooke, *Biotechnology* 1992, 10: 882-6). Apart from hybridization strategies, the application of ribozymes, i.e. RNA molecules that act as enzymes, destroying RNA that carries the message of disease has also been described (see e.g. Barinaga, *Science* 1993, 262: 1512-1514). In preferred embodiments, the subject to be treated is a human, and therapeutic antisense nucleic acids or derivatives thereof are directed against hTARPP. It is preferred that cells of the central nervous system, preferably the brain, of a subject are treated in such a way. Cell penetration can be performed by known strategies such as coupling of antisense nucleic acids and derivatives thereof to carrier particles, or the above described techniques. Strategies for administering targeted therapeutic oligodeoxynucleotides are known to those of skill in the art (see e.g. Wickstrom, *Trends Biotechnol* 1992, 10: 281-287). In some cases, delivery can be performed by mere topical application. Further approaches are directed to intracellular expression of antisense RNA. In this strategy, cells are transformed *ex vivo* with a recombinant gene that directs the synthesis of an RNA that is complementary to a region of target nucleic acid. Therapeutical use of intracellularly expressed antisense RNA is procedurally similar to gene therapy.

In further preferred embodiments, the method comprises grafting donor cells into the central nervous system, preferably the brain, of said subject, or donor cells preferably treated so as to minimize or reduce graft rejection, wherein said donor cells are genetically modified by insertion of at least one transgene encoding said agent or agents. Said transgene might be carried by a viral vector, in particular a retroviral vector. The transgene can be inserted into the donor cells by a nonviral physical transfection of DNA encoding a transgene,

in particular by microinjection. Insertion of the transgene can also be performed by electroporation; chemically mediated transfection, in particular calcium phosphate transfection, and liposomal mediated transfection.

In preferred embodiments, said agent for treating and preventing a neurodegenerative disease, in particular Alzheimer's disease, is a therapeutic protein which can be administered to said subject, preferably a human, by a process comprising introducing subject cells into said subject, said subject cells having been treated *in vitro* to insert a DNA segment encoding said therapeutic protein, said subject cells expressing *in vivo* in said subject a therapeutically effective amount of said therapeutic protein. Said DNA segment can be inserted into said cells *in vitro* by a viral vector, in particular a retroviral vector.

Methods of treatment, according to the present invention, comprise the application of therapeutic cloning, transplantation, and stem cell therapy using embryonic stem cells or embryonic germ cells and neuronal adult stem cells, combined with any of the previously described cell- and gene therapeutic methods. Stem cells may be totipotent or pluripotent. They may also be organ-specific. Strategies for repairing diseased and/or damaged brain cells or tissue comprise (i) taking donor cells from an adult tissue. Nuclei of those cells are transplanted into unfertilized egg cells from which the genetic material has been removed. Embryonic stem cells are isolated from the blastocyst stage of the cells which underwent somatic cell nuclear transfer. Use of differentiation factors then leads to a directed development of the stem cells to specialized cell types, preferably neuronal cells (Lanza et al., *Nature Medicine* 1999, 9: 975-977), or (ii) purifying adult stem cells, isolated from the central nervous system, or from bone marrow (mesenchymal stem cells), for *in vitro* expansion and subsequent grafting and transplantation, or (iii) directly inducing endogenous neural stem cells to proliferate, migrate, and differentiate into functional neurons (Peterson DA, *Curr. Opin. Pharmacol.* 2002, 2: 34-42). Adult neural stem cells are of great potential for repairing damaged or diseased brain tissues, as the germinal centers of the adult brain

are free of neuronal damage or dysfunction (Colman A, *Drug Discovery World* 2001, 7: 66-71).

In preferred embodiments, the subject for treatment or prevention, according to the present invention, can be a human, an experimental animal, e.g. a mouse or a rat, a domestic animal, or a non-human primate. The experimental animal can be an animal model for a neurodegenerative disorder, e.g. a transgenic mouse and/or a knock-out mouse with an Alzheimer's-type neuropathology.

In a further aspect, the invention features a modulator of an activity, or a level, or both said activity and said level of at least one substance which is selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii).

In an additional aspect, the invention features a pharmaceutical composition comprising said modulator and preferably a pharmaceutical carrier. Said carrier refers to a diluent, adjuvant, excipient, or vehicle with which the modulator is administered.

In a further aspect, the invention features a modulator of an activity, or a level, or both said activity and said level of at least one substance which is selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii) for use in a pharmaceutical composition.

In another aspect, the invention provides for the use of a modulator of an activity, or a level, or both said activity and said level of at least one substance which is selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP

and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii) for a preparation of a medicament for treating or preventing a neurodegenerative disease, in particular Alzheimer's disease.

In one aspect, the present invention also provides a kit comprising one or more containers filled with a therapeutically or prophylactically effective amount of said pharmaceutical composition.

In a further aspect, the invention features a recombinant, non-human animal comprising a non-native gene sequence coding for hTARPP, or a fragment, or a variant, or a derivative thereof. The generation of said recombinant, non-human animal comprises (i) providing a gene targeting construct containing said gene sequence and a selectable marker sequence, and (ii) introducing said targeting construct into a stem cell of a non-human animal, and (iii) introducing said non-human animal stem cell into a non-human embryo, and (iv) transplanting said embryo into a pseudopregnant non-human animal, and (v) allowing said embryo to develop to term, and (vi) identifying a genetically altered non-human animal whose genome comprises a modification of said gene sequence in both alleles, and (vii) breeding the genetically altered non-human animal of step (vi) to obtain a genetically altered non-human animal whose genome comprises a modification of said endogenous gene, wherein said gene is mis-expressed, or under-expressed, or over-expressed, and wherein said disruption or alteration results in said non-human animal exhibiting a predisposition to developing symptoms of neuropathology similar to a neurodegenerative disease, in particular Alzheimer's disease. Strategies and techniques for the generation and construction of such an animal are known to those of ordinary skill in the art (see e.g. Capecchi, *Science* 1989, 244: 1288-1292 and Hogan et al., 1994, *Manipulating the Mouse Embryo: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York). It is preferred to make use of such a recombinant non-human animal as an animal model for investigating neurodegenerative diseases, in particular Alzheimer's disease.

In another aspect, the invention features an assay for screening for a modulator of neurodegenerative diseases, in particular Alzheimer's disease, or related diseases and disorders of one or more substances selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii). This screening method comprises (a) contacting a cell with a test compound, and (b) measuring the activity, or the level, or both the activity and the level of one or more substances recited in (i) to (iv), and (c) measuring the activity, or the level, or both the activity and the level of said substances in a control cell not contacted with said test compound, and (d) comparing the levels of the substance in the cells of step (b) and (c), wherein an alteration in the activity and/or level of said substances in the contacted cells indicates that the test compound is a modulator of said diseases and disorders.

In one further aspect, the invention features a screening assay for a modulator of neurodegenerative diseases, in particular Alzheimer's disease, or related diseases and disorders of one or more substances selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii), comprising (a) administering a test compound to a test animal which is predisposed to developing or has already developed a neurodegenerative disease or related diseases or disorders, and (b) measuring the activity and/or level of one or more substances recited in (i) to (iv), and (c) measuring the activity and/or level of said substances in a matched control animal which is equally predisposed to developing or has already developed said diseases and to which animal no such test compound has been administered, and (d) comparing the activity and/or level of the substance in the animals of step (b) and (c), wherein an alteration in the activity and/or level of substances in the test animal indicates that the test compound is a modulator of said diseases and disorders.

In a preferred embodiment, said test animal and/or said control animal is a recombinant non-human animal which expresses hTARPP, or a fragment, or a variant thereof, or a derivative thereof, under the control of a transcriptional regulatory element which is not the native hTARPP gene transcriptional control regulatory element.

In another embodiment, the present invention provides a method for producing a medicament comprising the steps of (i) identifying a modulator of neurodegenerative diseases by a method of the aforementioned screening assays and (ii) admixing the modulator with a pharmaceutical carrier. However, said modulator may also be identifiable by other types of screening assays.

In another aspect, the present invention provides for an assay for testing a compound, preferably for screening a plurality of compounds, for inhibition of binding between a ligand and hTARPP, or a fragment or derivative thereof. Said screening assay comprises the steps of (i) adding a liquid suspension of said hTARPP, or a fragment or derivative thereof, to a plurality of containers, and (ii) adding a compound or a plurality of compounds to be screened for said inhibition to said plurality of containers, and (iii) adding fluorescently labelled ligand to said containers, and (iv) incubating said hTARPP, or said fragment or derivative thereof, and said compound or plurality of compounds, and said fluorescently labelled ligand, and (v) measuring the amounts of fluorescence associated with said hTARPP, or with said fragment or derivative thereof, and (vi) determining the degree of inhibition by one or more of said compounds of binding of said ligand to said hTARPP, or said fragment or derivative thereof. Instead of utilizing a fluorescently labelled ligand, it might in some aspects be preferred to use any other detectable label known to the person skilled in the art, e.g. radioactive labels, and detect it accordingly. Said method may be useful for the identification of novel compounds as well as for evaluating compounds which have been improved or otherwise



optimized in their ability to inhibit the binding of a ligand to a gene product of the gene coding for hTARPP, or a fragment or derivative thereof.

In one further embodiment, the present invention provides a method for producing a medicament comprising the steps of (i) identifying a compound as an inhibitor of binding between a ligand and a gene product of the gene coding for hTARPP by the aforementioned inhibitory binding assay and (ii) admixing the compound with a pharmaceutical carrier. However, said compound may also be identifiable by other types of screening assays.

In another aspect, the invention features an assay for testing a compound, preferably for screening a plurality of compounds to determine the degree of binding of said compounds to hTARPP, or to a fragment, a variant or derivative thereof. Said screening assay comprises (i) adding a liquid suspension of said hTARPP, or a fragment, or a variant, or derivative thereof, to a plurality of containers, and (ii) adding a fluorescently labelled compound or a plurality of fluorescently labelled compounds to be screened for said binding to said plurality of containers, and (iii) incubating said hTARPP, or said fragment, or variant, or derivative thereof, and said fluorescently labelled compound or fluorescently labelled compounds, and (iv) measuring the amounts of fluorescence associated with said hTARPP, or with said fragment, or variant, or derivative thereof, and (v) determining the degree of binding by one or more of said compounds to said hTARPP, or said fragment, or variant, or derivative thereof. In this type of assay it might be preferred to use a fluorescent label. However, any other type of detectable label might also be employed. Said method may be useful for the identification of novel compounds as well as for evaluating compounds which have been improved or otherwise optimized in their ability to bind to hTARPP.

In one further embodiment, the present invention provides a method for producing a medicament comprising the steps of (i) identifying a compound as a binder to a gene product of the gene coding for hTARPP by the aforementioned binding assays and (ii) admixing the compound with a

pharmaceutical carrier. However, said compound may also be identifiable by other types of screening assays.

In another embodiment, the present invention provides for a medicament obtainable by any of the methods according to the herein claimed screening assays. In one further embodiment, the instant invention provides for a medicament obtained by any of the methods according to the herein claimed screening assays.

Other features and advantages of the invention will be apparent from the following description of figures and examples which are illustrative only and not intended to limit the remainder of the disclosure in any way.

Figure 1 depicts the brain regions with selective vulnerability to neuronal loss and degeneration in Alzheimer's disease. Primarily, neurons within the inferior temporal lobe, the entorhinal cortex, the hippocampus, and the amygdala are subject to degenerative processes in Alzheimer's disease (Terry et al., *Annals of Neurology* 1981, 10:184-192). These brain regions are mostly involved in the processing of learning and memory functions. In contrast, neurons within the frontal cortex, the occipital cortex, and the cerebellum remain largely intact and preserved from neurodegenerative processes in Alzheimer's disease. Brain tissues from the frontal cortex (F) and the temporal cortex (T) of Alzheimer's disease patients and healthy, age-matched control individuals were used for the herein disclosed examples. For illustrative purposes, the image of a normal healthy brain was taken from a publication by Strange (*Brain Biochemistry and Brain Disorders*, Oxford University Press, Oxford, 1992, p.4).

Figure 2 illustrates the verification of the differential expression of the human TARPP (hTARPP) gene by quantitative RT-PCR analysis. Quantification of RT-PCR products from RNA samples collected from the frontal cortex (F) and temporal cortex (T) of Alzheimer's disease patients (Figure 2a) and of

healthy, age-matched control individuals (Figure 2b) was performed by the LightCycler rapid thermal cycling technique. The data were normalized to the combined average values of a set of standard genes which showed no significant differences in their gene expression levels. Said set of standard genes consisted of genes for cyclophilin B, the ribosomal protein S9, the transferrin receptor, GAPDH, and beta-actin. The figure depicts the kinetics of amplification by plotting the cycle number against the amount of amplified material as measured by its fluorescence. Note that the amplification kinetics of hTARPP cDNAs from both the frontal and temporal cortices of a normal control individual during the exponential phase of the reaction nearly overlap (Figure 2b, arrowheads), whereas in Alzheimer's disease (Figure 2a) there is a significant shift of the curve for the sample derived from frontal cortex, indicating a differential regulation of hTARPP mRNA expression in frontal cortex in comparison to temporal cortex.

Figure 3 discloses the protein sequence of human TARPP (hTARPP); SEQ ID NO. 1. The full length human TARPP protein consists of 814 amino acids.

Figure 4 shows an alignment of the amino acid sequence of SEQ ID NO.1, hTARPP protein, with mouse (*Mus musculus*) TARPP amino acid sequence (GenBank accession number af324451).

Figure 5 represents the nucleotide sequence of SEQ ID NO. 2, the coding sequence of the human TARPP gene, comprising 2442 nucleotides.

Figure 6 shows the nucleotide sequence of SEQ ID NO. 3, the hTARPP cDNA, comprising 3212 nucleotides. Primers used for quantitative PCR analysis are located from nucleotide 2471 to 2493 for the forward primer and from nucleotide 2518 to 2539 for the reverse primer.

Figure 7 depicts SEQ ID NO. 4, the nucleotide sequence of the 69 bp cDNA fragment, amplified with the primers used for quantitative PCR analysis. For the location of the primers refer to SEQ ID NO. 3, Figure 6.

Figure 8 charts the schematic alignment of SEQ ID NO. 4, the hTARPP cDNA fragment, SEQ ID NO. 2, the coding sequence of the hTARPP gene, and SEQ ID NO. 3, the hTARPP gene nucleotide sequence derived from the alignment of human EST nucleotide sequences found in the GenBank genetic sequence database. EST numbers are written on the left side, all sequences are 5' to 3' directed.

Table 1 lists expression levels in the frontal cortex relative to the temporal cortex for the transcription product of the hTARPP gene in six Alzheimer's disease patients (1.67-3.34  $\Delta$ -fold) and six healthy, age-matched control individuals (0.31-1.38  $\Delta$ -fold). The values shown are reciprocal values according to the formula described herein.

#### EXAMPLE I:

##### (i) Brain tissue dissection from patients with Alzheimer's disease:

Brain tissues from Alzheimer's disease patients and age-matched control subjects were collected within 6 hours post-mortem and immediately frozen on dry ice. Sample sections from each tissue were fixed in paraformaldehyde for histopathological confirmation of the diagnosis. Brain areas for differential expression analysis were identified (see Fig. 1) and stored at  $-80^{\circ}\text{C}$  until RNA extractions were performed.

##### (ii) Isolation of total RNA:

Total RNA was extracted from post-mortem brain tissue by using the RNeasy kit (Qiagen) according to the manufacturer's protocol. The accurate RNA concentration and the RNA quality was determined with the DNA LabChip system using the Agilent 2100 Bioanalyzer (Agilent Technologies). For additional quality testing of the prepared RNA, i.e. exclusion of partial degradation and testing for DNA contamination, specifically designed intronic GAPDH oligonucleotides and genomic DNA as reference control were utilised

to generate a melting curve with the LightCycler technology, as described in the supplied protocol by the manufacturer (Roche).

**(iii) Determination of differential expression by quantitative RT-PCR:**

In order to identify changes in gene expression in different tissues we examined differential expression of the hTARPP gene using the LightCycler technology (Roche). This technique features rapid thermal cycling for the polymerase chain reaction as well as real-time measurement of fluorescent signals during amplification and therefore allows for highly accurate quantification of RT-PCR products by using a kinetic rather than an endpoint approach. The ratio of hTARPP cDNA from the temporal cortex and frontal cortex was determined (relative quantification).

First, a standard curve was generated to determine the efficiency of the PCR with specific primers for hTARPP (5'-ACAGCCAATCATGCTACCTAACC-3' and 5'-ACAGTAAACAGGCATTCCAGTGG-3'). PCR amplification (95 °C and 1 sec, 56 °C and 5 sec, and 72 °C and 5 sec) was performed in a volume of 20 µl containing Lightcycler-FastStart DNA Master SYBR Green I mix (containing FastStart Taq DNA polymerase, reaction buffer, dNTP mix with dUTP instead of dTTP, SYBR Green I dye, and 1 mM MgCl<sub>2</sub>; Roche), 0.5 µM primers, 2 µl of a cDNA dilution series (final concentration of 40, 20, 10, 5, 1 and 0.5 ng human total brain cDNA; Clontech) and depending on the primers used, additional 3 mM MgCl<sub>2</sub>. Melting curve analysis revealed a single peak at approximately 81 °C with no visible primer dimers. Quality and size of the PCR product were determined with the DNA LabChip system (Agilent 2100 Bioanalyzer, Agilent Technologies). A single peak at the expected size of 69 bp for hTARPP was observed in the electropherogram of the sample.

In an analogous manner, the PCR protocol was applied to determine the PCR efficiency of a set of reference genes which were selected as a reference standard for quantification. In the present invention, the mean value of five such reference genes was determined: (1) cyclophilin B, using the specific primers 5'-ACTGAAGCACTACGGGCCTG-3' and 5'-AGCCGTTGGTGTCTT-TGCC-3' except for MgCl<sub>2</sub> (an additional 1 mM was added instead of 3 mM). Melting curve analysis revealed a single peak at approximately 87 °C with no

visible primer dimers. Agarose gel analysis of the PCR product showed one single band of the expected size (62 bp). (2) Ribosomal protein S9 (RPS9), using the specific primers 5'-GGTCAAATTTACCCTGGCCA-3' and 5'-TCTCATCAAGCGTCAGCAGTTC-3' (exception: additional 1 mM MgCl<sub>2</sub> was added instead of 3 mM). Melting curve analysis revealed a single peak at approximately 85°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (62 bp). (3) beta-actin, using the specific primers 5'-TGGAACGGTGAAGGTGACA-3' and 5'-GGCAAGGGACTTCTGTAA-3'. Melting curve analysis revealed a single peak at approximately 87°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (12 bp). (4) GAPDH, using the specific primers 5'-CGTCATGGGTG-TGAACCATG-3' and 5'-GCTAAGCAGTTGGTGGTGCAG-3'. Melting curve analysis revealed a single peak at approximately 83°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (81 bp). (5) Transferrin receptor TRR, using the specific primers 5'-GTCGCTGGTTCAGTTCGTGATT-3' and 5'-AGCAGTTGG-CTGTTGTACCTCTC-3'. Melting curve analysis revealed a single peak at approximately 83°C with no visible primer dimers. Agarose gel analysis of the PCR product showed one single band with the expected size (80 bp).

For calculation of the values, first the logarithm of the cDNA concentration was plotted against the threshold cycle number  $C_t$  for hTARPP and the five reference standard genes. The slopes and the intercepts of the standard curves (i.e. linear regressions) were calculated for all genes. In a second step, cDNAs from frontal cortex and temporal cortex were analyzed in parallel and normalized to cyclophilin B. The  $C_t$  values were measured and converted to ng total brain cDNA using the corresponding standard curves:

$$10^{((C_t \text{ value} - \text{intercept}) / \text{slope})} \quad [\text{ng total brain cDNA}]$$

The values for temporal cortex and frontal cortex cDNAs of hTARPP were normalized to cyclophilin B, and the ratio was calculated using the following formula:

$$\text{Ratio} = \frac{\text{hTARPP temporal [ng]} / \text{cyclophilin B temporal [ng]}}{\text{hTARPP frontal [ng]} / \text{cyclophilin B frontal [ng]}}$$

In a third step, the set of reference standard genes was analyzed in parallel to determine the mean average value of the temporal to frontal ratios of expression levels of the reference standard genes for each individual brain sample. As cyclophilin B was analyzed in step 2 and step 3, and the ratio from one gene to another gene remained constant in different runs, it was possible to normalize the values for hTARPP to the mean average value of the set of reference standard genes instead of normalizing to one single gene alone. The calculation was performed by dividing the ratio shown above by the deviation of cyclophilin B from the mean value of all housekeeping genes. The results of one such quantitative RT-PCR analysis for hTARPP are shown in Figure 2.

#### (v) Sequence Analysis

Searching the EST database of the GenBank database for sequence similarities to the identified differentially expressed human cDNA fragment (SEQ ID NO. 4), as stated in the present invention, it was found that SEQ ID NO. 4 is identical to portions of the human EST sequences hms80139 and bg201698 and others (shown in Figure 8). These human ESTs showed homology to mouse (*Mus musculus*) TARPP. Aligning human ESTs in addition to SEQ ID NO. 4, a complete EST cluster representing the hTARPP cDNA, SEQ ID NO. 3, was determined. The amino acid sequence of a large open reading frame, with the potential to encode a protein of 814 amino acid residues was deduced, SEQ ID NO. 1.

## CLAIMS

1. An isolated nucleic acid encoding a protein molecule shown in SEQ ID NO. 1.
2. An isolated nucleic acid molecule of claim 1, wherein the nucleic acid molecule is a D N A molecule.
3. An isolated nucleic acid molecule of claim 2, wherein the nucleic acid molecule is a cD N A molecule, in particular a cD N A molecule comprising a nucleotide sequence shown in SEQ ID NO. 2 or SEQ ID NO. 3.
4. An isolated D N A molecule capable of hybridizing with the complement of the cD N A described in SEQ ID NO. 2 or SEQ ID NO. 3 under stringent condition.
5. A vector comprising a nucleic acid molecule according to one of claims 1 to 4.
6. A vector according to claim 5 wherein said vector is a plasmid, a virus or a bacteriophage.
7. A cell transformed with a nucleic acid molecule according to one of claims 1 to 4, wherein said cell is in particular a bacterial cell, a yeast cell, a mammalian cell, or an insect cell.
8. A protein molecule shown in SEQ ID NO. 1.
9. An antibody specifically immunoreactive with an immunogen, wherein said immunogen is a protein molecule shown in SEQ ID NO. 1, or a fragment thereof.



10. Use of an antibody of claim 9, for detecting the pathological state of a cell in a sample from a subject, comprising immunocytochemical staining of said cell with said antibody, wherein an altered degree of staining, or an altered staining pattern in said cell compared to a cell representing a known health status indicates a pathological state of said cell, and wherein said pathological state relates to a neurodegenerative disease, preferably Alzheimer's disease.
11. A method of diagnosing or prognosticating a neurodegenerative disease in a subject, or determining whether a subject is at increased risk of developing said disease, comprising:  
determining a level and/or an activity of
- (i) a transcription product of the gene coding for hTARPP, and/or
  - (ii) a translation product of the gene coding for hTARPP, and/or
  - (iii) a fragment or derivative of said transcription or translation product,
- in a sample from said subject and comparing said level and/or said activity to a reference value representing a known disease or health status, thereby diagnosing or prognosticating said neurodegenerative disease in said subject, or determining whether said subject is at increased risk of developing said neurodegenerative disease.
12. A method of monitoring the progression of a neurodegenerative disease in a subject, comprising:  
determining a level and/or an activity of
- (i) a transcription product of the gene coding for hTARPP, and/or
  - (ii) a translation product of the gene coding for hTARPP, and/or
  - (iii) a fragment or derivative of said transcription or translation product,
- in a sample from said subject and comparing said level and/or said activity to a reference value representing a known disease or health status,

thereby monitoring the progression of said neurodegenerative disease in said subject.

13. A method of evaluating a treatment for a neurodegenerative disease, comprising:

determining a level and/or an activity of

- (i) a transcription product of the gene coding for hTARPP, and/or
- (ii) a translation product of the gene coding for hTARPP, and/or
- (iii) a fragment or derivative of said transcription or translation product,

in a sample from a subject being treated for said disease and comparing said level and/or said activity to a reference value representing a known disease or health status, thereby evaluating said treatment for said neurodegenerative disease.

14. The method according to any of claims 11 to 13 wherein said neurodegenerative disease is Alzheimer's disease.

15. The method according to any of claims 11 to 14 wherein said sample is a cell, or a tissue, or an organ, or a body fluid, in particular cerebrospinal fluid or blood.

16. The method according to any of claims 11 to 15 wherein said reference value is that of a level and/or an activity of

- (i) a transcription product of the gene coding for hTARPP, and/or
- (ii) a translation product of the gene coding for hTARPP, and/or
- (iii) a fragment or derivative of said transcription or translation product

in a sample from a subject not suffering from said neurodegenerative disease.

17. The method according to any of claims 11 to 16 wherein an increase or decrease in hTARPP RNA and/or hTARPP protein in a cell, or tissue, or

body fluid from said subject relative to a reference value representing a known health status indicates a diagnosis, or prognosis, or increased risk of Alzheimer's disease in said subject.

18. The method according to any of claims 11 to 17, further comprising comparing a level and/or an activity of

- (i) a transcription product of the gene coding for hTARPP, and/or
- (ii) a translation product of the gene coding for hTARPP, and/or
- (iii) a fragment or derivative of said transcription or translation product

in a series of samples taken from said subject over a period of time.

19. The method according to claim 18 wherein said subject receives a treatment prior to one or more of said sample gatherings.

20. The method according to claim 19 wherein said level and/or activity is determined before and after said treatment of said subject.

21. A kit for diagnosing or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, in a subject, or determining the propensity or predisposition of a subject to develop such a disease, said kit comprising:

- (a) at least one reagent which is selected from the group consisting of
  - (i) reagents that selectively detect a transcription product of the gene coding for hTARPP, and (ii) reagents that selectively detect a translation product of the gene coding for hTARPP, and
- (b) instruction for diagnosing, or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, or determining the propensity or predisposition of a subject to develop such a disease by
  - detecting a level, or an activity, or both said level and said activity, of said transcription product and/or said translation product of the gene coding for hTARPP, in a sample from said subject; and

- diagnosing or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, or determining the propensity or predisposition of said subject to develop such disease, wherein a varied level, or activity, or both said level and said activity, of said transcription product and/or said translation product compared to a reference value representing a known health status, or wherein a level, or activity, or both said level and said activity, of said transcription product and/or said translation product similar or equal to a reference value representing a known disease status indicates a diagnosis or prognosis of a neurodegenerative disease, in particular Alzheimer's disease, or an increased propensity or predisposition of developing such a disease.

22. A method of treating or preventing a neurodegenerative disease, in particular Alzheimer's disease, in a subject comprising administering to said subject in a therapeutically or prophylactically effective amount an agent or agents which directly or indirectly affect an activity and/or a level of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii).

23. A modulator of an activity and/or of a level of at least one substance which is selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii).

24. Use of a modulator of an activity and/or of a level of at least one substance which is selected from the group consisting of (i) a gene coding for hTARPP, and/or (ii) a transcription product of the gene coding for hTARPP, and/or (iii) a translation product of the gene coding for hTARPP, and/or (iv) a fragment or derivative of (i) to (iii) for a preparation of a

medicament for treating or preventing a neurodegenerative disease, in particular Alzheimer's disease.

25. A recombinant, non-human animal comprising a non-native gene sequence coding for hTARPP or a fragment thereof, or a derivative thereof, said animal being obtainable by:

- (i) providing a gene targeting construct comprising said gene sequence and a selectable marker sequence, and
- (ii) introducing said targeting construct into a stem cell of a non-human animal, and
- (iii) introducing said non-human animal stem cell into a non-human embryo, and
- (iv) transplanting said embryo into a pseudopregnant non-human animal, and
- (v) allowing said embryo to develop to term, and
- (vi) identifying a genetically altered non-human animal whose genome comprises a modification of said gene sequence in both alleles, and
- (vii) breeding the genetically altered non-human animal of step (vi) to obtain a genetically altered non-human animal whose genome comprises a modification of said endogenous gene, wherein said disruption results in said non-human animal exhibiting a predisposition to developing a neurodegenerative disease or related diseases or disorders.

26. An assay for screening for a modulator of neurodegenerative diseases, in particular Alzheimer's disease, or related diseases or disorders of one or more substances selected from the group consisting of

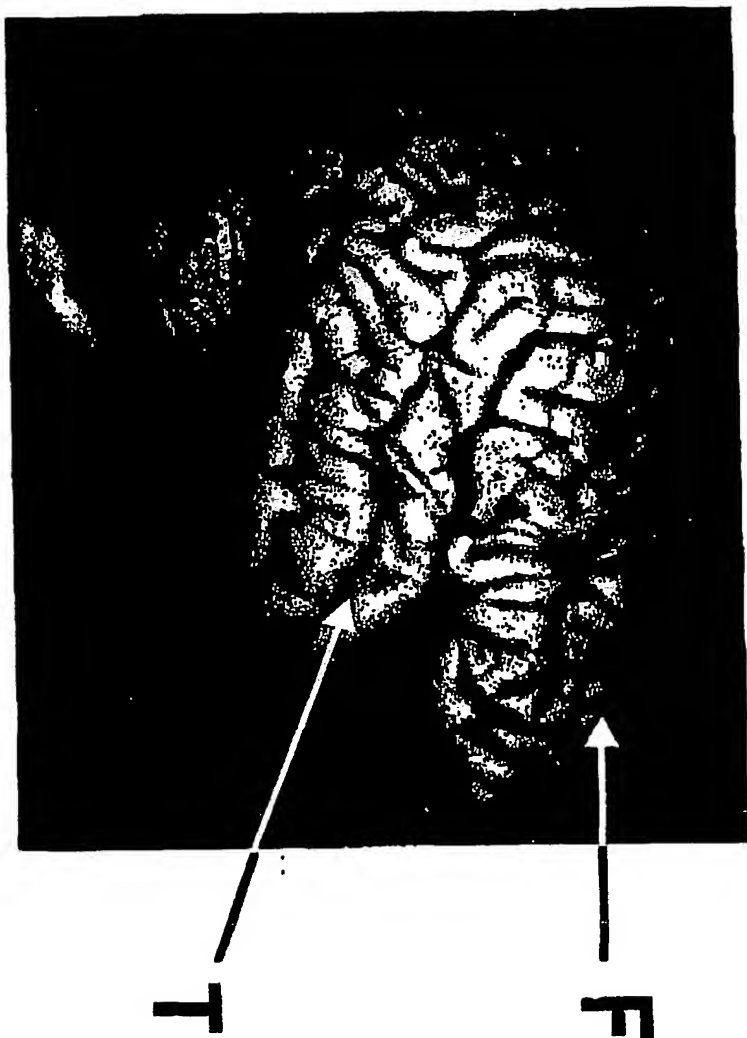
- (i) a gene coding for hTARPP, and/or
- (ii) a transcription product of the gene coding for hTARPP, and/or
- (iii) a translation product of the gene coding for hTARPP, and/or
- (iv) a fragment or derivative of (i) to (iii), said method comprising:
  - (a) contacting a cell with a test compound;

- (b) measuring the activity and/or level of one or more substances recited in (i) to (iv);**
- (c) measuring the activity and/or level of one or more substances recited in (i) to (iv) in a control cell not contacted with said test compound; and**
- comparing the levels and/or activities of the substance in the cells of step (b) and (c), wherein an alteration in the activity and/or level of substances in the contacted cells indicates that the test compound is a modulator of said diseases or disorders.**

**SUMMARY**

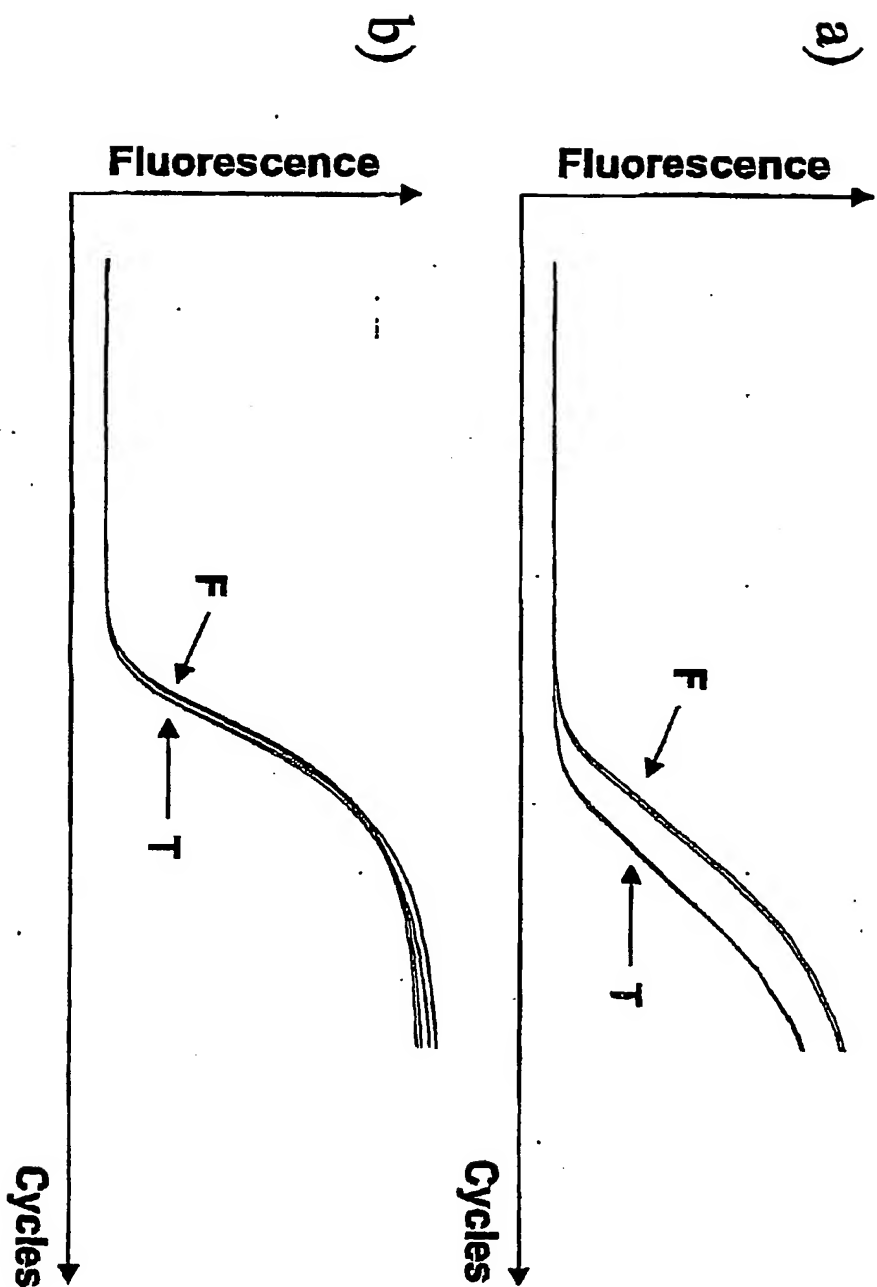
The present invention discloses a novel nucleic acid molecule encoding hTARPP. Further, the present invention discloses the differential expression of hTARPP in specific brain regions of Alzheimer's disease patients. Based on this finding, this invention provides a method for diagnosing or prognosticating a neurodegenerative disease, in particular Alzheimer's disease, or for determining whether a subject is at increased risk of developing a neurodegenerative disease, in particular Alzheimer's disease. Furthermore, this invention provides therapeutic and prophylactic methods for treating or preventing Alzheimer's disease and related neurodegenerative disorders using the gene coding for hTARPP. A method of screening for modulating agents of neurodegenerative diseases is also disclosed.

# Figure 1: Identification of Genes Involved in Alzheimer's Disease Pathology





**Figure 2: Verification of differential expression of human TARPP by quantitative RT-PCR**



## Figure 3 : SEQ ID NO. 1: amino acid sequence of human TARPP protein

Length: 814 aa

1 MSEQGDLNQA IAEEGGTBEQ TATPENGIVK SESLDDEEKL ELQRRLEAQN  
51 QERRKSKSGA GKGLTRSLA VCESSARPG GESLQDQESI HLQLSSFSSL  
101 QEEDKSRKDD SEREKEKDKN KDKTSEKPKI RMLSKDCSQE YTDSTGIDLH  
151 EFLINTLKNN SRDRMILLKM EQEIDFIAD NNNHYKKFPQ MSSYQRMLVH  
201 RVAAYFGLDH NVDQGTGKSVI INKTSSTRIP EQRFCHECLKD EKGBESQKRF  
251 ILKRDNSSID KEDNQSVCSQ ESLFVENSRL LEDSNICNET YKKRQLFRGN  
301 RDGSGRTSGS RQSSSENBK WSDHQRAWSS TSDSSNRNL KPAMTKTASF  
351 GGITVLTRGD STSSTRSTGK LSKAGSESSS SAGSSGSLSR THPPLQSTPL  
401 VSGVAAGSPG CVPYPENGIG GQVAPSSTSY ILLPLEAATG IPPGSILLNP  
451 HTGQPFVNPDP GTPAIYNPPT SQQPLRSAMV GQSQQQPPQQ QPSPQPPQQV  
501 QPPQPMAGP LVTQSVQGLQ ASSQSVQYPA VSFPQHLLP VSPTQHFFMR  
551 DDVATQFGQM TLRQSSGET PEPPSGPVYP SSLMPQPAQQ PSYVIASGQ  
601 QLPTGGFSGS GPPISQQVLQ PPPSPQGFVQ QPPAQMPVY YYPSGQYPTS  
651 TTQQYRPMAP VQYNAQR5QQ MPQAAQQAGY QPVL5GQQGF QGLIGVQQPP  
701 QSQNVINNQQ GTPVQ5VMV5 YPTMSSYQVP MTQGSQGLPQ QSYQQPIMLP  
751 NQAGQGSLPA TGM5V5CNVT PPTPQNNLRL IGP5CP5STV PVMSASCR5N  
801 CASMSNAGWQ VKF\*

**Figure 4: Alignment of SEQ ID NO. 1, human TARPP, with mouse TARPP**

**Length: 814 aa**

1	MSEQGDLNQAIAEEGGTEQETATPENGIVKSESLDEBEKLELQRRLEAQN	50
1	MSEQGGLTPTILEEBGQTEPESA.PENGILKSESLDEEEKLELQRRLEAQN	49
51	QERRKSKSGAGKGKLTRSLAVCEESSARPGGESLQDQESIHLQLSSFS	100
50	QERRKSKSGAGKGKLTRSLAVCEESSARSGGESHQDQESIHLQLSSFP	99
101	QEDDKSRKDDSEREKEKDKNKDKTSEKPKIRMLSKDCSQEYTDSTGID	150
100	QEDDKSRKDDSEREKEKDKNREKLSERPKIRMLSKDCSQEYTDSTGID	149
151	EFLINTLKNNSRDRMILLKMEQEIIDFIADNNNHKKFPQMSSYQRM	200
150	GFLINTLKNNSRDRMILLKMEQEMIDFIADSNNHKKFPQMSSYQRM	199
201	RVAAYFGLDHNVDQTGKSVIINKTSSTRIPEQRFCEHLKDEKGEESQ	250
200	RVAAYFGLDHNVDQTGKSVIINKTSSTRIPEQRFCEHLKDEKSEESQ	249
251	ILKRDNSSIDKEDNQ.....SV	267
250	ILKRDNSSIDKEDNQNRMHPPFRDDRKSIEEREYBYQVRERIFAHSV	299
268	CSQESLFVENSRLLEDNICNETYKKRQLFRGNRDGSGRTSGSRQSS	317
300	CSQESLFLDNSRLQEDMHICNETYKKRQLFRAHRDSSGRTSGSRQSS	349
318	ELKWSDHQRAWSSSTDSDSSNRNLKPAMTKTASFGGITVLTTRGDST	367
350	ELRWPDHQRAWSSSTDSDSSNRNLKPTMTKTASFGGITVLTTRGDST	399
368	TGKLSKAGSESSSSAGSSGSLSRTHPFLQSTPLVSGVAAGSPGCVPY	417
400	AGKLSKTGSESSSSAGSSGSLSRTHP..QSTALTSSVAAGSPGCMAY	447
418	GIGGQVAPSSSTSYILLPLEAATGIPPGSILLNPHTGQPFVNPDGTP	467
448	GMGGQVPPSSSTSYILLPLESATGIPPGSILLNPHTGQPFVNPDGTP	497
468	PPTSQQPLRSAMVQGSSQQQPPQQQPSQPQQQVQPPQPOMAGPLVTQ	517
498	PPGSQOTLRTVGGO..FOOPPPQQQPSQPQQQVOASQPOMAGPLVTQ	543

518 GLQASSQSVQYPAVSFPPQHLLPVSPTQHFFPMRDDVATQFGOMTLSRQSS 567  
|:-| || |:-| ||||  
544 .....REELAAQFSQLSMSRQSS 561  
568 GETPEPPSGPVYPSSLMPPAQQPSYVIASSTGQQLPTGGFSGSGPPISQQ 617  
|:||||| |||.||:| || |||| | |||| | |||| | |||| | ||||  
562 GDTPEPPSGTVYPASLLPQTAQFQSYVITSAGQQLSTCGFSDSGPPISQQ 611  
618 VLQPPPSPPQGFVQQPPPAQMFVYYYPSGQYPTSTTQQYRPMAPVQYNAQR 667  
||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||  
612 VLQAPPSPPQGFVQQPPPAQMSVYYYPSGQYPTSTSQQYRPLASVQYSAQR 661  
668 SQQMPQAAQQAGYQFVLSGQQGFQGLIGVQQPPQSONVINNQGTFFVQSV 717  
|||.|| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||  
662 SQQIPQTTQAGYQFVLSGQQGFQGMGVQQSAHSQGVMSQQGAPVHGV 711  
718 MVSYPMTSSYQVPMTQGSQGLPQQSYQQPIMLPNQAGQGSLLPATGMPVYC 767  
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||  
712 MVSYPMTSSYQVPMTQGSQAVPQQTYQPPIMLPNQAGQGSLLPATGMPVYC 761  
768 NVTPTFPQNNLRLIGPHCPSSSTVPVMSASCRINCASMSNAGWQVKF 813  
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||  
762 NVTPTFPQNNLRLMGPHCPSSSTVPVMSASCRINCASMSNAGWQVKF 807

## Figure 5: SEQ ID NO. 2: nucleotide sequence of human TARPP coding sequence

Length: 2442 bp

```

1  ATGTCTGAGC AAGGAGACCT GAATCAGGCA ATAGCAGAGG AAGGAGGGAC
51  TGAGCAGGAG ACGGCCACTC CAGAGAACGG CATTGTTAAA TCAGAAAGTC
101 TGGATGAAGA GGAGAAACTG GAACTCGAGA GCGCGCTGGA GGCTCAGAAAT
151 CAAGAAAGAA GAAAATCCAA GTCAGGAGCA GGAAAAGGTA AACTGACTCG
201 CAGYCTTGCT GTCTGTGAGG AATCTTCTGC CAGACCAGGA GGTGAAAGTC
251 TTCAGGATCA GGAATCAATT CATTACAGC TTTCCAGTTT TTCCAGCCTG
301 CAAGAGGAGG ATAAATCTAG GAAAGATGAC TCTGAAAGAG AAAAGAAAAA
351 GGATAAAAAC AAAGATAAAA CCTCTGAAAA ACCCAAGATC AGAATGTTAT
401 CAAAAGATTG CAGCCAAGAA TACACGGATT CTACAGGCAT AGACTTACAC
451 GAGTTTCTGA TTAACACATT AAAGAATAAT TCCAGGGACA GGATGATACT
501 TTTGAAAATG GASCAGGAAA TTATTGATTT CATTGCTGAC AACAAATAATC
551 ATTATAAAAA GTTCCCTCAG ATGTCATCGT ATCAGAGGAT GCTTGTCCTAT
601 CGAGTGGCAG CTTATTTTGG ATTGGATCAC AATGTGGATC AAACAGGAAA
651 ATCTGTTATC ATCAACAAGA CCAGCAGCAC CAGAATACCA GAGCAAGGTT
701 TTTGTGAACA TTTAAAAGAT GAAAAGGTTG AAGAATCCCA GAAGCGGTTT
751 ATCTTGAAGC GAGATAACTC TAGTATTGAT AAAGAAGACA ATCAGTCAGT
801 TTGCTCCAG GAAAGCCTTT TTGTGGAAAA CAGTAGGCTC TTGGAAGACA
851 GTAACATATG CAATGAGACC TATAAGAAAA GACAGCTCTT TCGGGGCAAC
901 AGAGATGGCT CAGGGAGAAC ATCTGGGAGT CGACAGAGCA GCTCAGAAAA
951 TGAACTCAAG TGGTCTGACC ACCAAAGGGC CTGGAGCAGC ACAGACTCCG
1001 ACAGTTCCAA CCGCAATCTA AAGCCCGCCA TGACCAAGAC GCGCAGTTTTT
1051 GGGGGCATCA CCGTGCTGAC CAGGGGTGAC AGCACTTCCA GTACTAGGAG
1101 TACCGGGAAG CTGTCCAAAG CAGGTTCCGA GTCTTCCAGC AGTGCAGGCT
1151 CCTCAGGATC GCTGTCCGCG ACCCATCCAC CTCTCCAGAG CACACCCCTA
1201 GTCTCAGGTG TGGCAGCTGG CTCTCCAGGC TGTGTGCCTT ATCCAGAGAA
1251 TGGAATAGGG GGCCAGGTTG CTCCAGCAG CACCAGCTAC ATCCTCCTTC
1301 CACTTGAAGC TGCAACAGGC ATCCCGCCTG GAAGCATCCT TCTTAATCCA
1351 CACACAGGCC AGCCCTTTGT GAATCCCGAT GGAACCTCTG CAATATACAA
1401 CCCACCCACC AGTCAGCAGC CCCTGCGAAG CGCCATGGTG GGGCAGTCCC
1451 AACACAGGCC GCCACAGCAG CAGCCCTCCC CGCAGCCCCA ACAGCAGGTC
1501 CAGCCACCGC AGCCACAGAT GGCAGGCCCT CTGGTCACTC AGTCTGTCCA
1551 GGGGCTGCAG GCTTCCTCCC AGTCAGTGCA ATATCCGGCA GTCTCTTTTC
1601 CTCCCCAGCA CCTCCTACCT GTGTCTCCAA CGCAGCACTT TCCCATGAGA
1651 GATGATGTGG CAACACAGTT TGGCCAGATG ACCCTGAGCC GGCAGTCCTC
1701 GGGGGAGACT CCTGAACCCC CATCAGGTCC TGTCTACCCA TCCTCCCTTA
1751 TGCCACAGCC GGCCAGCAG CCCAGCTATG TAATCGCCTC TACAGGCCAG
1801 CAGCTTCCCTA CAGGAGGATT CTCAGGCTCT GGCCTTCCA TCTCCAGCA
1851 GGTCTCCAG CCCCCCTCCT CACCACAGGG ATTCTGTCAA CAGCCTCCGC
1901 CTGCACAGAT GCCTGTATAT TATTACCCAT CTGGTCAGTA CCCTACCTCA
1951 ACCACGCAAC AGTACCGGCC CATGGCCCCG GTTCAGTACA ACGCTCAGAG
2001 GAGTCAACAG ATGCCACAGG CAGCACAGCA AGCAGGTTAC CAGCCAGTCT
2051 TGTCTGGTCA ACAGGGATTG CAAGGCCTAA TAGGAGTGCA GCAGCCACCT
2101 CAGAGTCAGA ACGTGATAAA TAACCAACAA GGAACCTCCG TGCAAAGCGT
2151 GATGGTTTCC TACCCAACAA TGTCTTCTTA TCAGGTGCCA ATGACCCAGG
2201 GTTCTCAAGG ACTGCCCCAG CAGTCATACC AACAGCCAAT CATGCTACCT

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NR. 6434 S: 51

2251 AACCAGGCAG GTCAAGGGTC ACTCCCAGCC ACTGGAATGC CTGTTTACTG  
2301 TAATGTCACA CCGCCCCACCC CTCAGAACAA CCTTAGGCTG ATTGGCCCCAC  
2351 ACTGCCCCCTC CAGCACTGTC CCAGTGATGT CAGCTAGCTG CAGAACAAAC  
2401 TGTGCAAGTA TGAGCAATGC TGGTTGSCAG GTCAAATTCT GA

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## Figure 6: SEQ ID NO. 3: nucleotide sequence of human TARPP cDNA

Length: 3212 bp

```
1  GTGATTGCT GGAAGCTGGT CATTAGTGTT GACGATGTGT CACACTGTGT
51  AAGGGAATCG CATGGAGATG GGCATTCCGA ACTGTTAATG GGGACATGGG
101 ACTCCAGTTG TCTCTGATCA CTTGTGTGGA TTTTCCTGGC GTAGAACGAC
151 AGAAGCCGCT AGTAAGTCGC CAAGACCTAC AGCAGGAATT CTGCACCAAA
201 GGGCATAAAA TCTTGTIATT TTAATTTGCA TCTGGGAGAA TGTCTGAGCA
251 AGGAGACCTG AATCAGGCAA TAGCAGAGGA AGGAGGGACT GAGCAGGAGA
301 CGGCCACTCC AGAGAACGGC ATTGTTAAAT CAGAAAGTCT GGATGAAGAG
351 GAGAAACTGG AACTGCAGAG GCGGCTGGAG GCTCAGAATC AAGAAAGAAG
401 AAAATCCAAG TCAGGAGCAG GAAAAGGTAA ACTGACTCGC AGYCTTGCTG
451 TCTGTGAGGA ATCTTCTGCC AGACCAGGAG GTGAAAGTCT TCAGGATCAG
501 GAATCAATTC ATTTACAGCT TTCCAGTTTT TCCAGCCTGC AAGAGGAGGA
551 TAAATCTAGG AAAGATGACT CTGAAAGAGA AAAAGAAAAG GATAAAAACA
601 AAGATAAAAC CTCTGAAAAA CCAAGATCA GAATGTTATC AAAAGATTGC
651 AGCCAAGAAT ACACGGATTG TACAGGCATA GACTTACACG AGTTTCTGAT
701 TAACACATTA AAGAATAATT CCAGGGACAG GATGATACCT TTGAAAATGG
751 AGCAGGAAAT TATTGATTTC ATTGCTGACA ACAATAATCA TTATAAAAAG
801 TTCCCTCAGA TGTCATCGTA TCAGAGGATG CTGTCCATC GAGTGGCAGC
851 TTATTTTGGA TTGGATCACA ATGTGGATCA AACAGGAAAA TCTGTTATCA
901 TCAACAAGAC CAGCAGCACC AGAATACCAG AGCAAAGGTT TTGTGAACAT
951 TTAAAGATG AAAAAGGTGA AGAATCCCAG AAGCGGTTTA TCTTGAAGCG
1001 AGATAACTCT AGTATTGATA AAGAAGACAA TCAGTCAGTT TGCTCCCAGG
1051 AAAGCCTTTT TGTGAAAAAC AGTAGGCTCT TGGAAACACG TAACATATGC
1101 AATGAGAGCT ATAAGAAAAG ACAGCTCTTT CGGGGCAACA GAGATGGCTC
1151 AGGGAGAACA TCTGGGAGTC GACAGAGCAG CTGAGAAAAT GAACTCAAGT
1201 GGTCTGACCA CCAAGGGGCC TGGAGCAGCA CAGACTCCGA CAGTTCCAAC
1251 CGCAATCTAA AGCCCGCCAT GACCAAGACC GCGAGTTTTG GGGGCATCAC
1301 GGTGCTGACC AGGGGTGACA GCACTTCCAG TACTAGGAGT ACCGGGAAGC
1351 TGTCCAAAGC AGGTTCCGAG TCTTCCAGCA GTGCAGGCTC CTCAGGATCG
1401 CTGTCCCGCA CCCATCCACC TCTCCAGAGC ACACCCCTAG TCTCAGGTGT
1451 GGCAGCTGGC TCTCCAGGCT GTGTGCCTTA TCCAGAGAAT GGAATAGGGG
1501 GCCAGGTTGC TCCCAGCAGC ACCAGCTACA TCCTCCTTCC ACTTGAAGCT
1551 GCAACAGGCA TCCCGCCTGG AAGCATCCTT CTTAATCCAC ACACAGGCCA
1601 GCCCTTTGTG AATCCCGATG GAACTCCTGC AATATACAAC CCACCCACCA
1651 GTCAGCAGCC CCTGCGAAGC GCCATGGTGG GGCAGTCCCA ACAGCAGCCG
1701 CCACAGCAGC AGCCCTCCCC GCAGCCCCAA CAGCAGGTCC AGCCACCGCA
1751 GCCACAGATG GCAGGCCCTC TGGTCACTCA GTCTGTCCAG GGGCTGCAGG
1801 CTTCTTCCCA GTCAGTGCAA TATCCGCGAG TCTCTTTTCC TCCCCAGCAC
1851 CTCCTACCTG TGTCTCCAAC GCAGCACTTT CCCATGAGAG ATGATGTGGC
1901 AACACAGTTT GGCCAGATGA CCCTGAGCCG GCAGTCTTCG GGGGAGACTC
1951 CTGAACCCCC ATCAGGTCTT GTCTACCCAT CCTCCCTTAT GCCACAGCCG
2001 GCCCAGCAGC CCAGCTATGT AATCGCCTCT ACAGGCCAGC AGCTTCCTAC
2051 AGGAGGATTC TCAGGCTCTG GCCCTCCCAT CTCCAGCAGG GTCTTCCAGC
2101 CCCCTCCCTC ACCACAGGGA TTCGTGCAAC AGCCTCCGCC TGCACAGATG
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2201 GTACCGGCCC ATGGCCCCGG TTCAGTACAA CGTCCAGAGG AGTCAACAGA
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2301 CAGGGATTCC AAGGCCTAAT AGGAGTGCAG CAGCCACCTC AGAGTCAGAA  
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2401 ACCCAACAAT GTCTTCTTAT CAGGTGCCAA TGACCCAGGG TTCTCAAGGA  
2451 CTGCCCCAGC AGTCATACCA ACAGCCAATC ATGCTACCTA ACCAGGCAGG  
2501 TCAAGGGTCA CTCCCAGCCA CTGGAATGCC TGTTTACTGT AATGTCACAC  
2551 CGCCCAACCC TCAGAACAAC CTTAGGCTGA TTGGCCCACA CTGCCCCCTC  
2601 AGCACTGTCC CAGTGATGTC AGCTAGCTGC AGAACAACT GTGCAAGTAT  
2651 GAGCAATGCT GGTGGCAGG TCAAATTCG AGAGCTCTGG CTGTGGTACA  
2701 TTTCTTCAGA TATTTCTCAT GGCCTTTGAT GGAAGAGGAA CAAGGTGGGA  
2751 AAAGTGGCTG AGGACTTAAG TATTCACCTA ACACCTCAAAT GATTGCTGCT  
2801 GGTATTCTGT AAAAAGTAAA CAAAGACTAA TATACACGTT AGCTGGTTAA  
2851 TGGTGATAT TCTGTCATG TCTGCTAGGT ATGCCTTTAT AGCTTAGCTA  
2901 GTGACATGAA TTCATCAAGG TAAGATTCTC TCCTACCACT GAATACCACT  
2951 GTGTAGATTA TAATATCCCT AATTTGGATT AGTTTTGTAC TTTGTGTTGA  
3001 GTTTGTGATG CTAAAAGTAT TTAATAATTA TATACTAAAT CACATTGTAC  
3051 CAAAGCTGTA ATGGAAAAGC AAAGAAGAAC TGATGAATTG AAGGAATAAT  
3101 TTATATACAT TATAGAGTTT TCTTTTAA TGGATATATA CTGTATTGTA  
3151 GTGTTTAATC AAAATAAAAC TATTTGACCT TATGGAGGAA GGTGATGTTT  
3201 TTACCACTAA AA



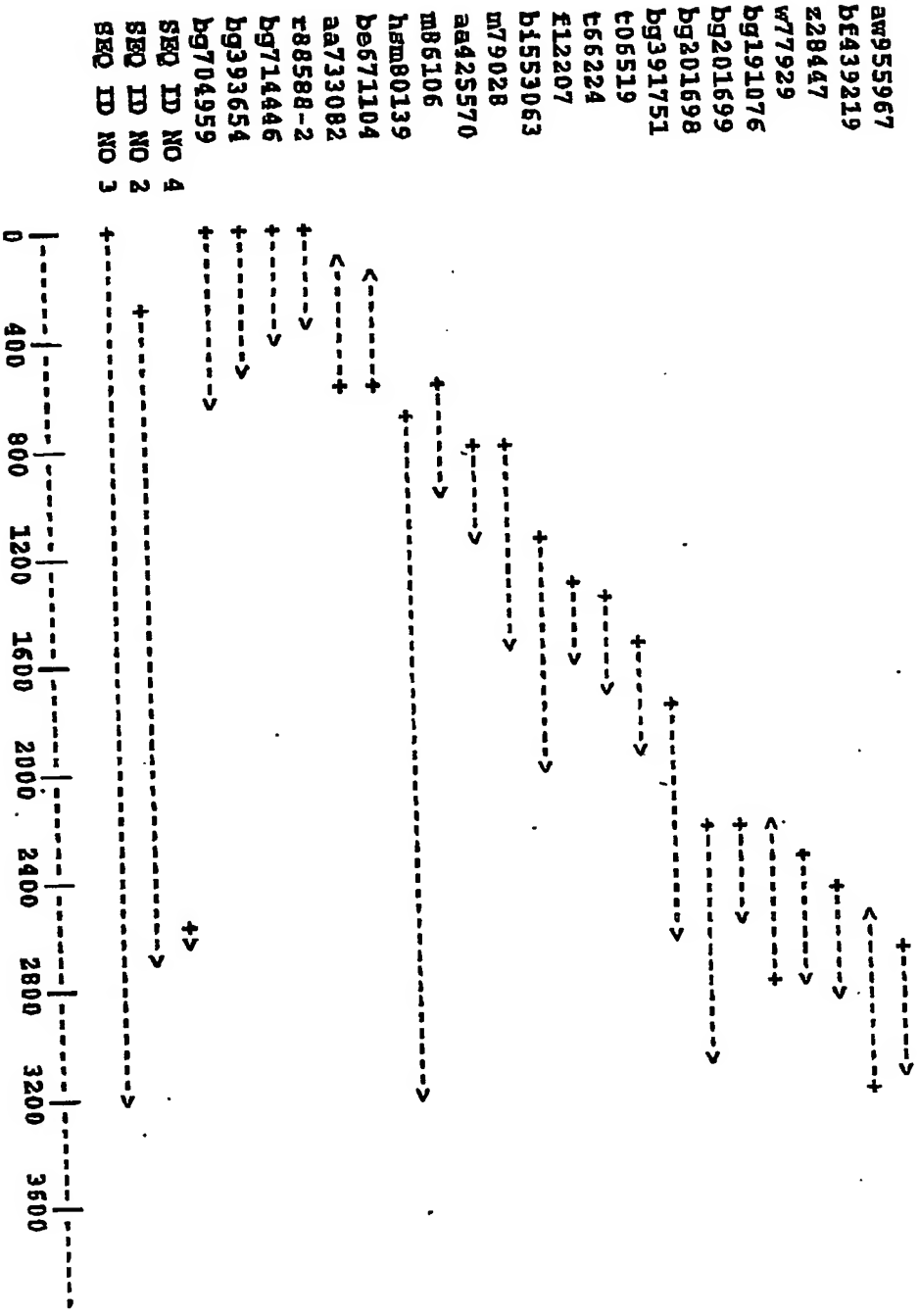
7 / 9

## Figure 7: SEQ ID NO. 4

Length: 69 bp

2471 ACAGCCAATCATGCTACCTAACCAGGCAGGTCAAGGGTCACTCCCAGCCA  
CTGGAATGCCTGTTTACTG 2539

**Figure 8: Schematic alignment of SEQ ID NO. 3, SEQ ID NO. 2 and SEQ ID NO. 4 with Genome Database EST-cluster**



**Table 1:**

<b>sample</b>	<b><math>\Delta</math> (fold)</b> <b>(frontal / temporal cortex)</b>
<b>patient 1</b>	<b>2.60</b>
<b>patient 2</b>	<b>2.74</b>
<b>patient 3</b>	<b>3.34</b>
<b>patient 4</b>	<b>2.21</b>
<b>patient 5</b>	<b>1.67</b>
<b>patient 6</b>	<b>1.82</b>
<b>control 1</b>	<b>0.67</b>
<b>control 2</b>	<b>0.63</b>
<b>control 3</b>	<b>1.05</b>
<b>control 4</b>	<b>1.38</b>
<b>control 5</b>	<b>0.31</b>
<b>control 6</b>	<b>0.63</b>